

Dynamic conformational ensembles: challenges of multiple time scales and multidomain proteins



Zoltán Gáspári, Nóra Epresi

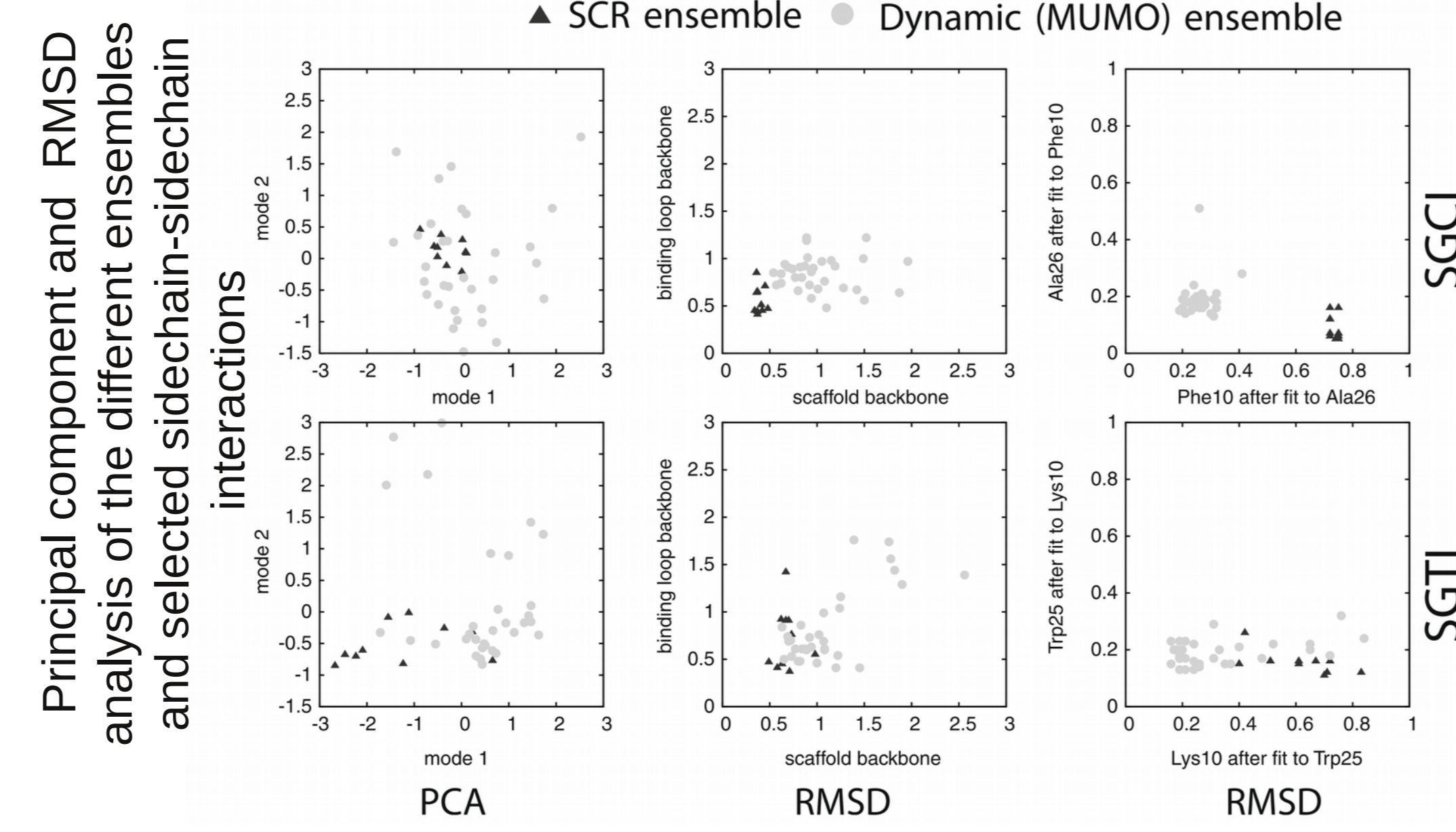
Faculty of Information Technology and Bionics,
Pázmány Péter Catholic University, Budapest, Hungary

<http://itk.ppke.hu/~gaszo> | gaspari.zoltan@itk.ppke.hu



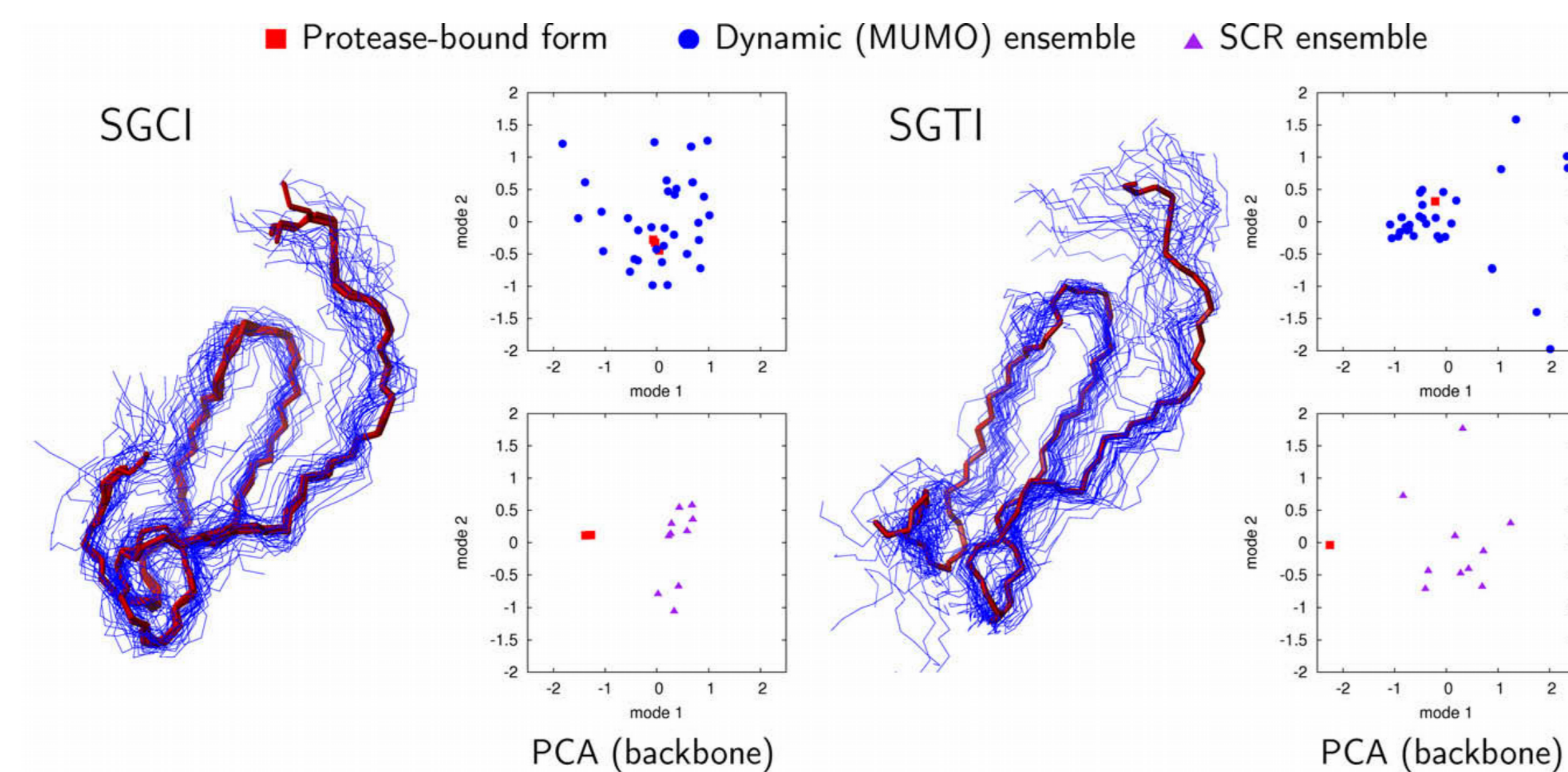
A previous application: small canonical serine protease inhibitors

- Standard mechanism, canonical inhibitors comprise multiple, unrelated protein families
- Substrate-like interaction with the target enzyme
- Common structural motif: protease binding loop
- Conventional view of efficiency [1]:
 - Rigidity of the binding loop is key
 - No conformational change upon enzyme binding
 - Based primarily on X-ray structures of free and complexed inhibitors
- NMR studies consistently reveal increased flexibility in the protease binding loop (ps-ns time scale)
- SGCI & SGTI: small inhibitors from desert locust with high efficiency and dynamic overall structure



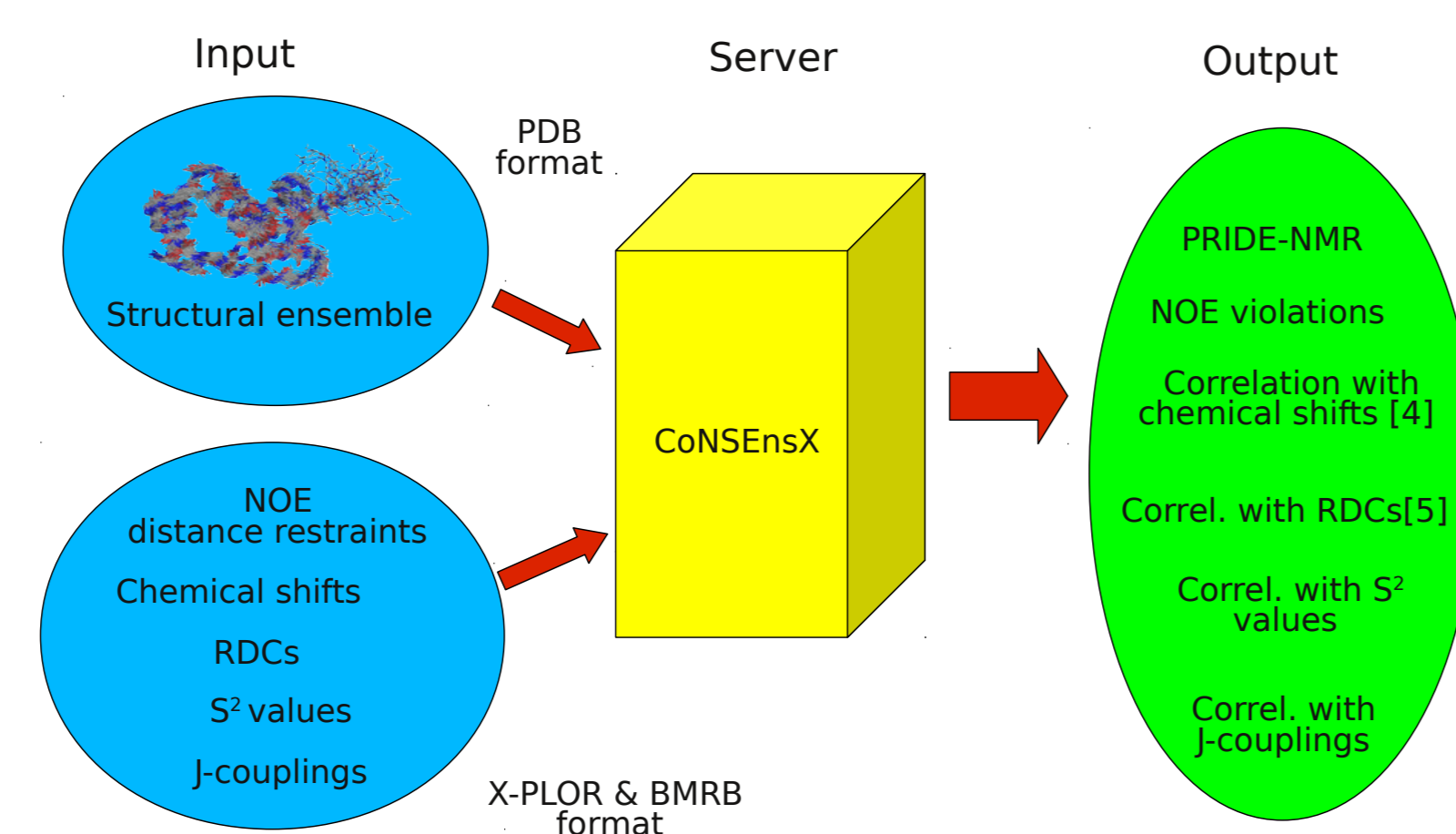
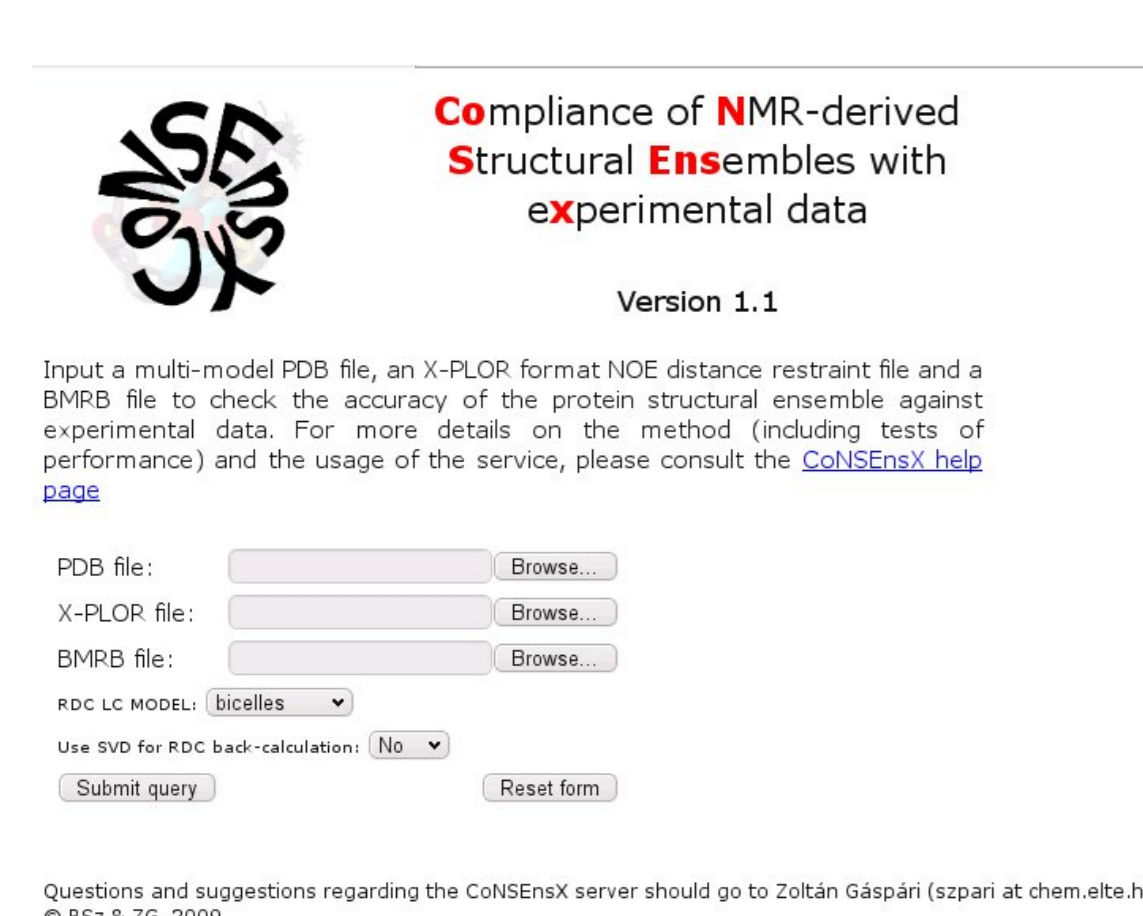
- Dynamic structural ensembles of the inhibitors are more diverse than SCR (single-conformer refinement) ones
- At the same time, they are more realistic than those calculated by conventional methods

Principal component analysis of the free and complexed structures of the inhibitors using different structural ensembles



- Enzyme-bound conformers are present in the solution state
- The conformational transitions are much faster (ps-ns scale) than the association with the enzyme (ms time scale)
- No energetic cost of protease binding related to conformational changes in 'nanosecond-timescale conformer selection'
- This is in agreement with the expectations of the rigid binding loop model [2]

The CoNSEnsX web server to analyze dynamic conformational ensembles



- <http://consensx.chem.elte.hu> [3]
- Assesses the correspondence of back-calculated observables to measured ones for the full ensemble

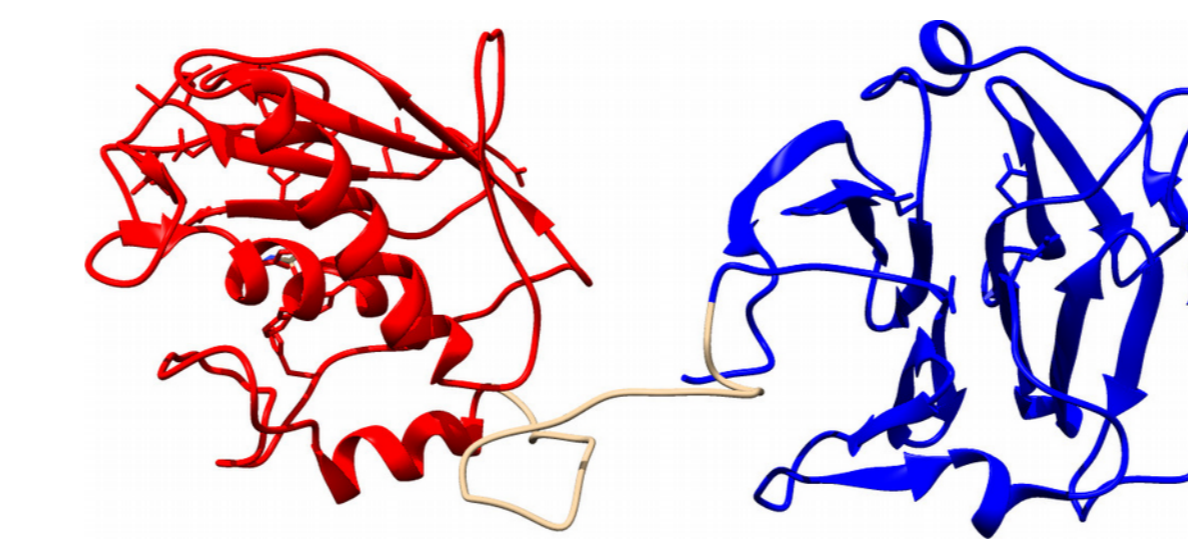
References

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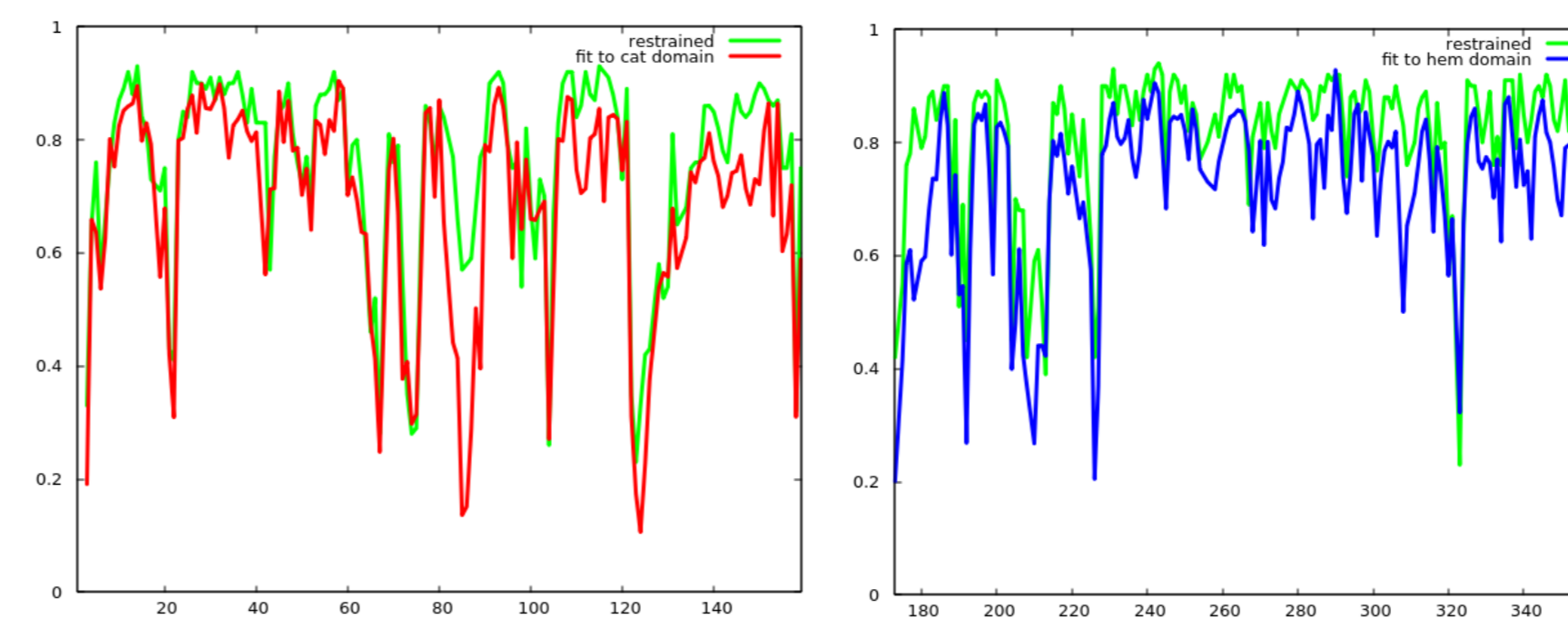
Details of the implementation

- In-house extensions for GROMACS 3.3.1 & 4.5.5, available for download on our home page
- Usage conforms to GROMACS conventions, restraints can be specified in the topology, other parameters in the run control file
- Order parameters (S^2) implemented as described in [6] and with fitting to reference group at each calculation step
 - Modification #1: (only in 3.3.1 yet): an individual fit group can be defined for not fully rigid molecules
 - Modification #2: can be defined for sub-ensembles for the generation of ensembles reflecting motions on multiple time scales
- NOE: pairwise averaging over replicas is implemented (similarly to the MUMO approach [7])

Exploratory calculations



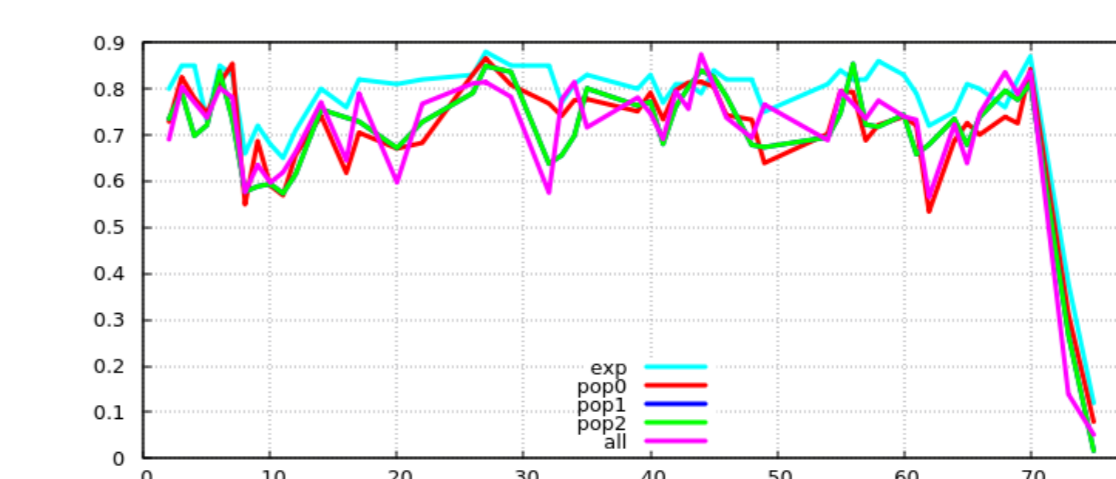
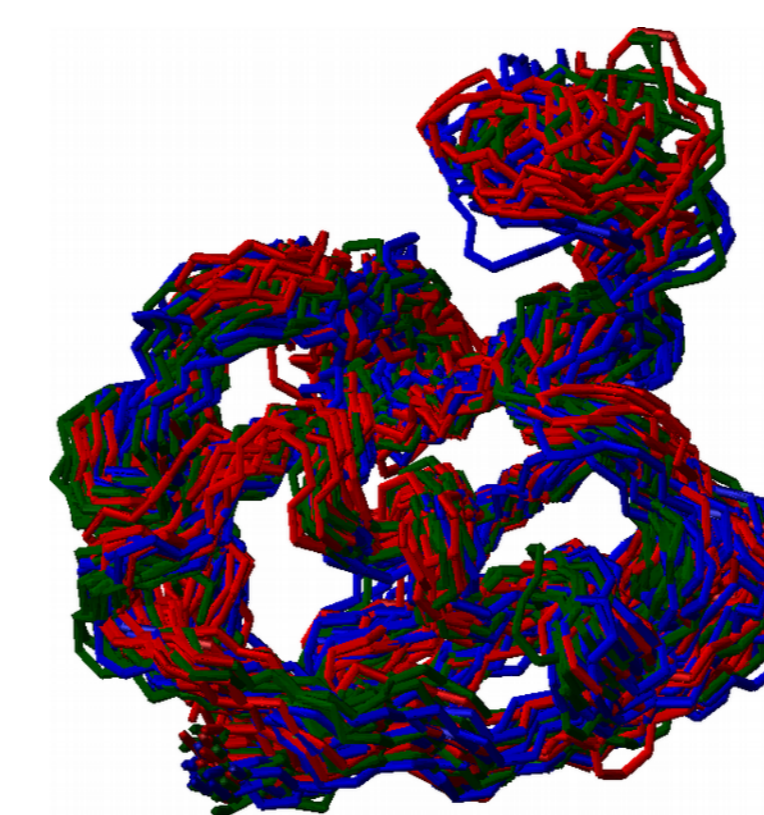
- The catalytic (red) and hemopexin-like (blue) domains of MMP12 show largely independent reorientation [7]
- Using synthetic S^2 values back-calculated from a 5 ns MD run (GROMACS 4.5.5, amber99sb FF, TIP3P water) for both domains separately
- Restraining in GROMACS 3.3.1. (0.3 ns, oplaa FF, 4 replicas) with two fit groups corresponding to the two domains



Challenges:

- Get better agreement for the flexible loops
- Can we obtain meaningful S^2 values from experimental relaxation data for such a protein?

- Ubiquitin displays motions on the submicrosecond time scale that are not present on the ps-ns time scale [9]
- Using experimental S^2 values and back-calculated RDCs from the EROS ensemble (PDB ID 3K39)
- Restraining in GROMACS 4.5.5. (1.2 ns, amber99sb FF, 3X4 replicas)



| | N-H S^2 | | N-H RDC | | C α -C RDC | | | | |
|------|---------------|-------|---------------|-------|-------------------|------|-------|-------|------|
| | Pearson Q (%) | RMSD | Pearson Q (%) | RMSD | Pearson Q (%) | RMSD | | | |
| pop0 | 0.936 | 10.81 | 0.08 | 0.905 | 51.06 | 2.66 | 0.872 | 68.8 | 0.74 |
| pop1 | 0.927 | 12.15 | 0.09 | 0.979 | 45.93 | 2.12 | 0.957 | 61.76 | 0.60 |
| pop2 | 0.896 | 12.94 | 0.09 | 0.972 | 39.53 | 1.95 | 0.960 | 49.74 | 0.53 |
| all | 0.917 | 14.93 | 0.10 | 0.975 | 41.87 | 2.01 | 0.944 | 59.07 | 0.60 |

Challenges:

- Find the optimal combination of relative restraint weights as well as the number of replicas and sub-ensembles
- Find the motional modes that discriminate the motions on different time scales
- Try to understand the connections between motions by detailed analysis of residue-residue interactions

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